

Fat Mass Localization Alters Fuel Oxidation during Exercise in Normal Weight Women

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ABSTRACT

ISACCO, L., P. DUCHE, D. THIVEL, A. MEDDAHI-PELLE, S. LEMOINE-MOREL, M. DUCLOS, and N. BOISSEAU. Fat Mass Localization Alters Fuel Oxidation during Exercise in Normal Weight Women. *Med. Sci. Sports Exerc.*, Vol. 45, No. 10, pp. 1887–1896, 2013. **Purpose:** Abdominal and lower body fat mass tissues exhibit particular metabolic profiles at rest and during exercise. However, data are missing in normal weight women during exercise. The purpose of this study was to investigate the effect of low (LA/LB) and high (HA/LB) abdominal to lower body (A/LB) fat mass ratio on metabolic and hormonal responses during exercise in premenopausal normal weight women. **Methods:** After preliminary testing ($\dot{V}O_{2\max}$ and body composition assessment), substrate oxidation (RER, lipid, and carbohydrate oxidation rates), metabolic response (glycerol, free fatty acids, and glucose), and hormonal response (insulin, growth hormone, atrial natriuretic peptide, adrenaline, and noradrenaline) were determined during exercise (45 min at 65% of $\dot{V}O_{2\max}$) in 21 premenopausal normal weight women (10 HA/LB women vs 11 LA/LB women). **Results:** Waist circumference was significantly higher in HA/LB women compared with LA/LB women ($P < 0.01$). No difference in other anthropometric characteristics, $\dot{V}O_{2\max}$, and resting blood values was observed between the two groups. LA/LB subjects exhibited greater lipid oxidation rates compared with HA/LB women during exercise ($P < 0.01$). This occurred with lower plasma insulin ($P < 0.05$) and glucose ($P < 0.05$) concentrations and higher plasma free fatty acids ($P < 0.05$), glycerol ($P < 0.05$), growth hormone ($P < 0.05$), and atrial natriuretic peptide levels ($P < 0.01$) during exercise in the LA/LB group compared with the HA/LB group. **Conclusions:** The present study demonstrated that LA/LB women exhibited an increase in whole-body lipid mobilization and use during exercise compared with HA/LB counterparts. This greater reliance on lipid as fuel metabolism during exercise could be explained by substrate availability and metabolic and hormonal responses. It appeared that LA/LB women exhibited greater metabolic flexibility during an exercise bout of 45 min at 65% of $\dot{V}O_{2\max}$ on cycle ergometer. **Key Words:** ADIPOSITY PHENOTYPE, ENERGY METABOLISM, HORMONES, FEMALE POPULATION

Physical activity is one of the primary factors supporting good health and weight management because it favors an increase in lipolysis and promotes fat oxidation (32). In healthy normal weight subjects, sexual dimorphism is observed concerning substrate oxidation during exercise. Compared with men, women exhibit greater reliance on fat oxidation at the same relative exercise

intensity (34). Different factors such as specific estrogen levels (14,15), free fatty acids (FFA) availability (7), sex difference in catecholamines stimulated lipolysis (5), or total amount of fat mass (FM) and adipose tissue distribution (16,40) may explain these sex differences. In premenopausal women, a higher proportion of lower body FM (mainly subcutaneous tissue) is observed whereas abdominal FM (mainly visceral) is more developed in men (5,24). Abdominal fat depot is preferentially associated with metabolic disorders, such as insulin resistance, dyslipidemia, and metabolic syndrome, whereas lower body FM has been proposed as a protective factor against metabolic disruptions (9,30,33). From a metabolic point of view, there are some evidences showing that individuals with more abdominal FM than lower body FM display substrate mobilization and utilization impairments (5,18). Specifically, in the female population, from menopause, women exhibit greater abdominal FM (due to lower estrogen levels) and a decline in total lipid oxidation at rest and during exercise (35,36). Kanaley et al. (18) observed a greater FFA availability

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during exercise in lower body obese women compared with abdominal obese women without any difference in total lipid oxidation rates. A decrease in abdominal to lower body FM distribution (waist to hip ratio) in obese women was associated with higher FFA use at rest (20) and after a low-intensity exercise training program (40% $\dot{V}O_{2\max}$, three times per week for 12 wk) (38). If changes in insulin concentrations and insulin and adrenergic sensitivities may in part explain such differences (17), scientific evidences remain limited so far to explain these adaptations.

Several techniques have been used to determine body FM localization, such as selected anthropometric variables (e.g., waist to hip ratio), dual-energy x-ray absorptiometry (DXA), and other imaging techniques (e.g., magnetic resonance imaging) (4,8,18,20,33). Today, DXA is commonly used and offers a precise estimation of total and localized body composition as it is well correlated with magnetic resonance imaging results (4,8,29).

Fat mass distribution in postmenopausal or obese women may induce changes in substrate oxidation at rest or during exercise. However, to the best of our knowledge, there is no data available on how FM distribution affects substrate oxidation in normal weight premenopausal women. Thus, the purpose of the present study was to investigate the influence of abdominal to lower body (A/LB) FM ratio in normal weight premenopausal women on substrate oxidation at rest and during a moderate-intensity exercise. On the basis of sexual dimorphism and data obtained on postmenopausal and obese women, we hypothesized that normal weight premenopausal women, with lower A/LB FM ratio, would exhibit greater reliance on lipid as fuel metabolism during exercise compared with women with higher A/LB FM ratio.

METHODS

Participants

A total of 21 recreationally active women (mean \pm SEM age = 22.0 ± 0.59 yr; <4 h of physical activity per week, determined by interviews) were recruited by posted notices and e-mail. None were pregnant, and all had been weight stable at least for 3 months before the start of the experimentation. All

subjects were premenopausal with normal weight (body mass index [BMI] values within the healthy weight range ($19.5 < \text{BMI} < 25 \text{ kg}\cdot\text{m}^{-2}$) and waist circumference ($\text{WC} \leq 80 \text{ cm}$ [2]). As no standard exists concerning A/LB FM ratio in premenopausal lean women, ratios were calculated for the whole population ($n = 21$, 0.80 ± 0.03 , ranged from 0.56 to 1.06), and according to the median (0.78), women were divided in two groups: low A/LB FM ratio when the ratio was lower than 0.78 (LA/LB: $n = 11$, 0.68 ± 0.02 , ranged from 0.56 to 0.77) and high A/LB FM ratio when the ratio was higher than 0.78 (HA/LB: $n = 10$, 0.90 ± 0.03 , ranged from 0.82 to 1.06). As we had an odd number, the sample of both groups could not be equal. To assess the adequacy of the resulting classification, we performed a discriminant analysis introducing lipid oxidation rates during exercise as the main outcome. The analysis confirmed the two group classification.

Among the 21 participants, 11 were oral contraceptive users (low-dose monophasic combined oral contraception; ethinyl estradiol $\leq 30 \mu\text{g}$). Ten women were eumenorrheic with regular menstruations (length of cycles = 28 ± 0.5 d for at least 1 yr). Five eumenorrheic women and six oral contraceptive users were part of the LA/LB group, and five eumenorrheic women and five oral contraceptive users were part of the HA/LB group. For better standardization, eumenorrheic women were all tested during their luteal phase, whereas oral contraceptive users participated during the active phase of pill consumption. On the basis of clinical and biochemical findings, none of them had hirsutism or polycystic ovarian syndrome. The characteristics of all participants are presented in Table 1. Informed consent form was obtained from each subject, and the study protocol was approved by the relevant ethical authorities (CPP Sud Est VI-AU818) and complied with the Declaration of Helsinki.

Experimental Design

All women attended the laboratory on three separate occasions. During the first visit, an initial screening interview and a physical examination including anthropometric measurements and body composition assessment were performed before including participants to the study. A second preliminary

TABLE 1. Subject characteristics.

Variables	LA/LB ($n = 11$)	HA/LB ($n = 10$)	P
Age (yr)	22.5 ± 1.16	21.6 ± 0.39	NS
Height (m)	1.65 ± 0.01	1.66 ± 0.01	NS
Weight (kg)	61.13 ± 1.50	63.59 ± 2.19	NS
BMI ($\text{kg}\cdot\text{m}^{-2}$)	21.31 ± 0.62	23.38 ± 0.74	NS
WC (cm)	70.36 ± 1.07	77.00 ± 2.09	**
%FM	25.61 ± 1.33	27.78 ± 0.10	NS
FFM (kg)	41.44 ± 0.64	42.46 ± 1.69	NS
$\dot{V}O_{2\max}$ ($\text{mL}\cdot\text{min}^{-1}\cdot\text{kg}^{-1}$ FFM)	53.28 ± 1.80	49.79 ± 2.40	NS
A/LB FM ratio	0.68 ± 0.02	0.90 ± 0.03	***
A/LB FFM ratio	0.16 ± 0.02	0.17 ± 0.01	NS

Data are presented as mean \pm SEM.

** $P < 0.01$ and *** $P < 0.001$.

NS, no significant difference between groups; A/LB, abdominal to lower body FM ratio; LA/LB, low abdominal to lower body FM ratio group; HA/LB, high abdominal to lower body FM ratio group; BMI, body mass index; WC, waist circumference; FM, fat mass; FFM, fat-free mass; $\dot{V}O_{2\max}$, maximal oxygen consumption.

session was then arranged to familiarize individuals with the experimental procedures and to determine their maximal oxygen consumption ($\dot{V}O_{2\max}$). In a third experimental session, participants were asked to complete a 45-min cycle test (from 12:00 p.m. to 12:45 p.m.) 3 h after consumption of individually standardized breakfast. On the third session, blood samples (15 mL) were drawn prior, during and at the end of exercise (8:00 a.m., 12:00 p.m., 12:30 p.m., and 12:45 p.m.).

Anthropometric and body composition measurements. A digital scale was used to measure body mass to the nearest 0.1 kg, and barefoot standing height was assessed to the nearest 0.1 cm by using a wall-mounted stadiometer. BMI was calculated as body mass (kg) divided by height squared (m^2). WC was measured in a standing position with a non-elastic tape that was applied horizontally midway between the costal arch and the iliac crest. Body composition (fat-free mass [FFM] and FM) was determined by DXA (fan beam DXA, QDR 4500 x-ray bone densitometer; Hologic, Bedford, MA).

Adipose and FFM tissue localization—abdominal to lower body (A/LB) FFM and FM ratios. From DXA analysis, abdominal FM (visceral and subcutaneous tissues) were determined manually by an experienced technician by drawing a rectangular box around the region of interest between vertebral bodies L1 and L4. The upper limit was set with the horizontal line going through the T12/L1 vertebral space, and the lowest limit was set with a horizontal line going through the L4 and L5 vertebral space (4,13). Data were analyzed with Hologic QDR software for Windows (version 12.6), which integrates whole-body measurements and standard body regions, such as the trunk, arms, and lower limbs delineated by specific anatomical landmarks. Lower body FM (subcutaneous tissue) were also assessed from lower limb measurement.

The A/LB FM ratio was calculated as follows:

$$\text{A/LB FM ratio} = \frac{\text{abdominal fat mass (g)}}{\text{lower body fat mass (g)}}$$

Similarly, A/LB FFM ratio was calculated from FFM located in the abdominal region of interest and lower body FFM. The A/LB FFM ratio was calculated to account for differences between groups for FFM localization.

Preliminary visit—maximal exercise testing. $\dot{V}O_{2\max}$ was measured during a graded exhaustive exercise test on a cycle ergometer (Ergoline, Bitz, Germany). After a 4-min warm-up at 75 W, power output was increased by 25 W increments every 3 min until participant's exhaustion (test lasted between 10 and 15 min after warm-up). Participants were strongly encouraged by the experimenters throughout the test to perform a maximal effort. Respiratory gases ($\dot{V}O_2$ and $\dot{V}CO_2$) were measured breath-by-breath through a mask connected to O_2 and CO_2 analyzers (Oxycon pro-Delta, Jaeger, Hoechberg, Germany). $\dot{V}O_{2\max}$ was determined as the highest oxygen uptake for a 15-s period. Ventilatory parameters were averaged every 30 s. ECG was monitored throughout the test.

The criteria, which have been adopted to assess the achievement of $\dot{V}O_{2\max}$, included the following: 1) maximal heart rate within 10% of age-predicted maximal values ($220 - \text{age} \pm 10$ beats per minute); 2) RER values higher than 1.1; and 3) oxygen uptake reaching a plateau with increasing work rate.

Experimental session. Participants attended the laboratory at 7:30 a.m. They were asked to avoid alcohol and any food containing biogenic amines during 24 h preceding the test, which can alter the catecholamine analysis. Subjects were also asked to avoid any kind of strenuous exercise the day before the experimental session.

Individually standardized breakfast. Three hours before the start of the exercise testing, subjects received an individually standardized breakfast at 8:15 a.m., which had to be consumed within the next 45 min. Energy content represented $39.75 \text{ kJ} \cdot \text{kg}^{-1}$ body mass, and qualitative aspects were respected (55% CHO [$21.86 \text{ kJ} \cdot \text{kg}^{-1}$; $1.31 \text{ g} \cdot \text{kg}^{-1}$], 30% lipid [$11.93 \text{ kJ} \cdot \text{kg}^{-1}$; $0.32 \text{ g} \cdot \text{kg}^{-1}$], and 15% protein [$5.96 \text{ kJ} \cdot \text{kg}^{-1}$; $0.36 \text{ g} \cdot \text{kg}^{-1}$]). Breakfast included milk, sugar, bread, butter, and fruit. Coffee and chocolate were not allowed.

In addition, during the week before experimental session, eating habit interviews were realized to control spontaneous subjects' energy intake. All women exhibited well-balanced energy consumption and no difference appeared between the two groups (mean: $7949 \text{ kJ} \cdot \text{d}^{-1}$; 53% CHO [$67.95 \text{ kJ} \cdot \text{kg}^{-1}$; $4.06 \text{ g} \cdot \text{kg}^{-1}$], 32% lipid [$41.05 \text{ kJ} \cdot \text{kg}^{-1}$; $1.09 \text{ g} \cdot \text{kg}^{-1}$], and 15% protein [$19.25 \text{ kJ} \cdot \text{kg}^{-1}$; $1.15 \text{ g} \cdot \text{kg}^{-1}$]). They also confirmed that the investigated week was representative of their current diet.

Exercise test. Participants performed a 45-min exercise bout on a cycle ergometer at 65% of their $\dot{V}O_{2\max}$. The test began 3 h after the end of breakfast, approximately at 12:00 p.m. For each individual, the workload during the session was adjusted throughout the exercise bout to keep $\dot{V}O_2$ constant at 65% of $\dot{V}O_{2\max}$. Respiratory gases ($\dot{V}O_2$ and $\dot{V}CO_2$) were measured breath by breath through a mask connected to O_2 and CO_2 analyzers (Oxycon pro-Delta, Jaeger, Hoechberg, Germany). Ventilatory parameters were averaged every 30 s. ECG was monitored throughout the test. RER ($\dot{V}CO_2/\dot{V}O_2$) was calculated at rest (12:00 p.m.) then at the 30th (12:30 p.m.) and the 45th minute (end of exercise: 12:45 p.m.) of exercise. Fat and carbohydrate (CHO) oxidation rates were calculated from $\dot{V}O_2$ and $\dot{V}CO_2$ measurements according to Peronnet and Massicotte (31) equations:

$$\begin{aligned}\text{CHO}(\text{mg} \cdot \text{min}^{-1}) &= 4.585\dot{V}CO_2(\text{mL} \cdot \text{min}^{-1}) - 3.2255\dot{V}O_2(\text{mL} \cdot \text{min}^{-1}) \\ \text{Fat}(\text{mg} \cdot \text{min}^{-1}) &= 1.6946\dot{V}O_2(\text{mL} \cdot \text{min}^{-1}) - 1.7012\dot{V}CO_2(\text{mL} \cdot \text{min}^{-1})\end{aligned}$$

Fat and CHO oxidation rates were expressed as milligrams per minute per kilogram of FFM and calculated at rest (12:00 p.m.), at the 30th minute (12:30 p.m.), at and the 45th minute (12:45 p.m.) of exercise. Energy expenditure (kJ) during exercise was calculated as follows: $\dot{V}O_2 (\text{L} \cdot \text{min}^{-1}) \times \text{energy equivalent of oxygen} \times 45$ (duration, min).

Blood samples. When arriving at the laboratory (7:30 a.m.), a venous catheter was inserted into an antebrachial vein. Participants had to sit quietly for 30 min, and then the first blood sample was collected (8:00 a.m.). Samples were first collected in fasting condition (8:00 a.m.) to determine subjects' fasting biologic profile, then right before exercise (12:00 p.m.), at the 30th minute of exercise (12:30 p.m.), and at the end of exercise (45th minute of exercise: 12:45 p.m.). Catecholamines were measured only before (12:00 p.m.) and at the end of exercise (12:45 p.m.).

At every blood collection, hematocrit was immediately measured in duplicates by microcentrifugation (Sigma 1–14). Samples were centrifuged (4000g for 10 min at 4°C), aliquoted, and stored at –80°C until analysis.

Biochemical Assays

Plasma triglycerides (TG), total cholesterol, HDL-C, LDL-C, glycerol, FFA, and glucose were assessed using an automated analyzer (Konelab 20; Thermo Electron, Waltham, MA). The biochemical assay kits were purchased from Randox Laboratories (Crumlin, UK). Plasma insulin, growth hormone (GH), testosterone, and atrial natriuretic peptide (ANP) were measured by enzyme-linked immunosorbent assay with Euromedex kits (Paris, France). Plasma adrenaline and noradrenaline were determined by high-performance liquid chromatography, following the method of Koubi et al. (19) and using Euromedex kits (Paris, France).

All blood analyses were performed at the same time. Detection limits of TG, total cholesterol, HDL-C, glycerol, FFA, glucose, insulin, ANP, GH, testosterone, and catecholamines were 0.02 mmol·L⁻¹, 0.1 mmol·L⁻¹, 0.04 mmol·L⁻¹, 0.07 g·L⁻¹, 0.140 mEq·L⁻¹, 0.1 mmol·L⁻¹, 28 pg·mL⁻¹, 5.5 pg·mL⁻¹, 0.06 ng·mL⁻¹, 0.3 nmol·L⁻¹, and 0.06 nmol·mL⁻¹, respectively. The intra-assay coefficients of TG, total cholesterol, HDL-C, glycerol, FFA, glucose, insulin, ANP, GH, testosterone, and catecholamines were 1%, 1.1%, 0.8%, 3.9%, 1.5%, 1.8%, 2.3%, 2.7%, 2.3%, 4.9%, 5.3%, and 4.5%, respectively.

As plasma volume changes may occur during acute exercise, all metabolic and hormonal concentrations were corrected according to plasma volume fluctuations from hematocrit changes as proposed by Van Beaumont (39).

Calculations

Metabolic and hormonal responses during exercise were also expressed as the area under the response curve (AUC) calculated with trapezoid integration. Two indices of insulin resistance were calculated from glucose and insulin concentrations: glucose–insulin ratio (G/I) and Homeostatic Model Assessment–Insulin resistance (HOMA-IR) index: (fasting insulin level × fasting glucose level) / 22.5. From WC and TG values, we determined whether women exhibited hypertriglyceridemic waist (HTGW). HTGW is characterized by a WC ≥ 88 cm and TG concentrations ≥ 150 mg·dL⁻¹ (22).

Statistical Analysis

All statistical analyses were carried out with Statistica software (version 8.00, USA). On the basis of previous results measuring CHO oxidation during exercise in women (37), sample size estimation was performed before the beginning of the protocol to ensure a statistical power >90% (considering alpha level = 0.05, SD = 1 mg·min⁻¹·kg⁻¹, and minimal difference between groups from 1.5 mg·min⁻¹·kg⁻¹).

Results are expressed as mean ± SEM. The normality of the distribution was tested with the Kolmogorov–Smirnov test, and the homogeneity of variance was tested with the *F*-test. To assess the adequacy of the group classification, we performed a discriminant analysis introducing lipid oxidation rates during exercise as the main outcome. The analysis confirmed the two group classification. Physiological and anthropometric characteristics of the subjects were compared between groups with unpaired *t*-tests. The effect of group and time for all other variables was assessed by using a one-way (factor: group) ANOVA with repeated measures (time). When a significant effect was found, *post hoc* multiple comparisons were made by Newman–Keuls. Pearson correlations were used to test relationships between variables. Statistical significance was set up at *P* < 0.05.

RESULTS

Participant Characteristics

Total body mass, BMI, %FM, FFM, and A/LB FFM ratio were not significantly different between groups (*P* = 0.57, 0.53, 0.21, 0.58, and 0.13, respectively), whereas significant A/LB FM ratio characterized each group (*P* = 0.0005) (Table 1). WC was higher in the HA/LB group compared with the LA/LB group (*P* = 0.009) (Table 1). Physical fitness level, assessed by $\dot{V}O_{2\max}$ values, was not significantly different between the two groups (*P* = 0.34). Biological profiles in the fasting condition were similar between the LA/LB and the HA/LB groups, and no woman exhibited HTGW (Table 2).

Metabolic and Hormonal Responses to Exercise

There was no significant difference (*P* = 0.59) in energy expenditure during exercise between the LA/LB group (1236 ± 18 kJ) and the HA/LB group (1223 ± 21 kJ).

Substrate Use

RER. RER values were significantly greater in the HA/LB group compared with the LA/LB group during the exercise session (*P* = 0.005), but no time effect was observed (*P* = 0.11).

Substrate oxidation (mg·min⁻¹·kg⁻¹ FFM). At rest, no difference was observed in CHO and lipid oxidation rates between the two groups (*P* = 0.10 and 0.11, respectively). During exercise, CHO oxidation rates were lower (*P* = 0.01) and lipid oxidation rates were higher (*P* = 0.002) in the LA/LB group compared with the HA/LB group (Fig. 1). A

TABLE 2. Biological profiles in fasting condition in both groups.

Variables	LA/LB (n = 11)	HA/LB (n = 10)	P
HDL-C (mmol·L ⁻¹)	4.22 ± 0.22	3.73 ± 0.22	NS
LDL-C (mmol·L ⁻¹)	3.24 ± 0.26	3.52 ± 0.28	NS
Total cholesterol (mmol·L ⁻¹)	5.23 ± 0.28	5.10 ± 0.21	NS
TG (mmol·L ⁻¹)	1.11 ± 0.11	0.97 ± 0.10	NS
Glucose (mmol·L ⁻¹)	5.22 ± 0.11	5.38 ± 0.11	NS
Insulin (pmol·L ⁻¹)	24.10 ± 1.25	19.52 ± 1.25	NS
G/I	0.82 ± 0.18	0.68 ± 0.17	NS
HOMA-IR	0.82 ± 0.18	0.84 ± 0.31	NS
Testosterone (nmol·L ⁻¹)	6.11 ± 1.25	6.87 ± 1.49	NS
HTGW	No	No	

Data are presented as mean ± SEM.

NS, no significant difference between groups for all variables; LA/LB, low abdominal to lower body FM ratio group; HA/LB, high abdominal to lower body FM ratio group; HDL-C, high-density lipoprotein cholesterol; LDL, low-density lipoprotein cholesterol; HTGW, hypertriglyceridemic waist.

time effect was also observed for CHO ($P < 10^{-6}$) and lipid ($P < 10^{-6}$). A group–time interaction indicated that the LA/LB group had higher lipid oxidation rates at 30 and 45 min of exercise ($P = 0.008$) compared with the HA/LB group (Fig. 1B). A significant correlation was observed between the lipid oxidation rates and the A/LB FM ratio ($r = -0.51$, $P = 0.01$), but no significant correlation was observed between the CHO oxidation rates and the ratio ($r = 0.24$, $P = 0.25$). Lipid provided 21.34% and 12.15% of the total energy expenditure during exercise in LA/LB and HA/LB women, respectively.

Glucose and insulin responses. Under resting conditions and in fed state (12:00 p.m., before exercise), plasma glucose concentrations were not different between the two groups ($P = 0.10$), whereas plasma insulin concentrations were lower in LA/LB compared with HA/LB ($P = 0.0009$). G/I at 12:00 p.m. demonstrated a significant group difference as LA/LB women exhibited greater values than HA/LB (0.19 ± 0.03 and 0.09 ± 0.03 , respectively, $P = 0.04$),

indicating a lower insulin sensitivity in HA/LB for the same glycemia.

Exercise induced a decrease in plasma glucose levels in both groups (time effect: $P = 0.02$). Plasma glucose values were lower in LA/LB compared with HA/LB (group effect: $P = 0.03$) (Fig. 2A). During exercise, glucose AUC was also lower in the LA/LB group than that in the HA/LB group ($P = 0.0006$) (Fig. 2B).

Insulin concentrations decreased during exercise in both groups ($P < 10^{-6}$). A significant difference was observed between groups ($P = 0.02$) with greater insulin values in HA/LB. A group–time interaction showed that the LA/LB group had lower insulin concentrations at rest and at 30 min of exercise ($P = 0.001$) compared with HA/LB (Fig. 2C). During exercise, AUC for insulin was higher in HA/LB than that in LA/LB women ($P = 0.0004$) (Fig. 2D). No significant correlation was observed for insulin ($r = 0.16$, $P = 0.49$), glucose ($r = 0.29$, $P = 0.20$) concentrations, and A/LB FM ratio.

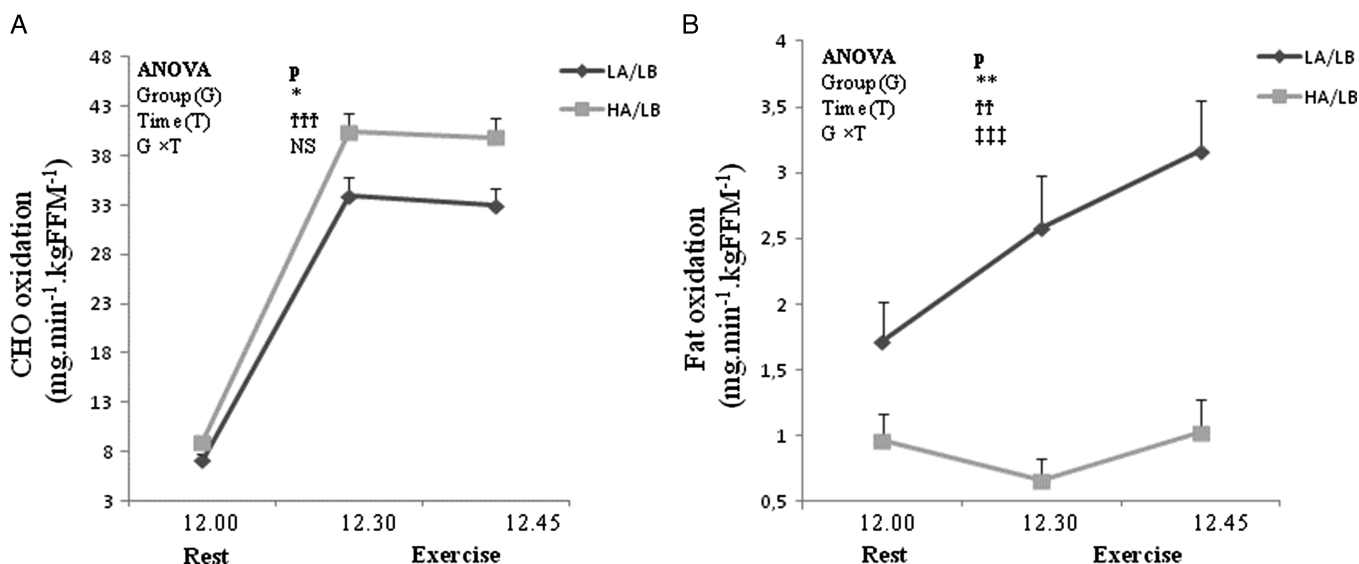


FIGURE 1—Substrate oxidation rates at rest and during exercise in both groups. A, CHO oxidation rates at rest and during exercise in both groups. B, Fat oxidation rates at rest and during exercise in both groups. Data are presented as mean ± SEM. NS, not statistically significant; group effect, $*P < 0.05$ and $**P < 0.01$; time effect, $††P < 0.01$ and $†††P < 0.001$; interaction group × time, $††††P < 0.001$. LA/LB, low abdominal to lower body FM ratio group; HA/LB, high abdominal to lower body FM ratio group; CHO, carbohydrates.

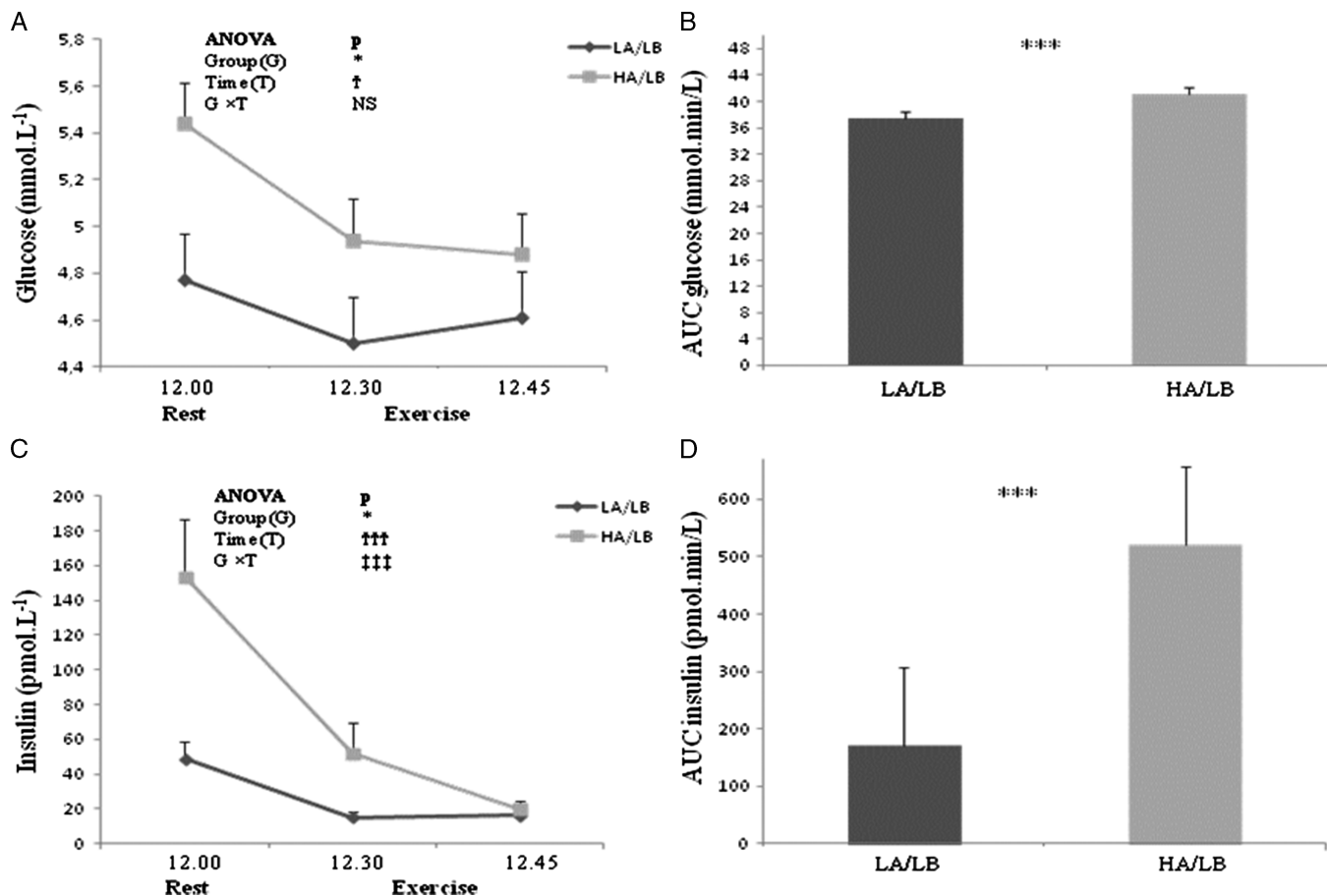


FIGURE 2—Glucose and insulin responses at rest and during exercise in both groups. A, Glucose responses at rest and during exercise in both groups. B, AUC for glucose in both groups. C, Insulin responses at rest and during exercise in both groups. D, AUC for insulin in both groups. Data are presented as mean \pm SEM. NS, not statistically significant; group effect, $*P < 0.05$ and $***P < 0.001$; time effect, $\dagger P < 0.05$ and $\dagger\dagger\dagger P < 0.001$; interaction group \times time, $\dagger\dagger\dagger P < 0.001$. LA/LB, low abdominal to lower body FM ratio group; HA/LB, high abdominal to lower body FM ratio group.

FFA and glycerol responses. Plasma FFA and glycerol concentrations were not different between the two groups at 12:00 p.m. ($P = 0.79$ and 0.33 , respectively). Exercise induced an increase in plasma FFA concentrations in all subjects (time effect: $P = 0.003$), with lower values in HA/LB compared with LA/LB (group effect: $P = 0.03$) (Fig. 3A). A group–time interaction demonstrated that LA/LB presented higher FFA levels at the 30th and 45th minute of exercise ($P = 0.006$). During exercise, AUC for FFA was higher in LA/LB compared with HA/LB women ($P = 0.0006$) (Fig. 3B). A significant correlation was found between FFA concentrations and A/LB FM ratio ($r = -0.47$, $P = 0.03$).

Glycerol concentrations increased during exercise (time effect: $P = 10^{-6}$) in both groups and were higher in LA/LB than that in HA/LB (group effect: $P = 0.03$) (Fig. 3C). Glycerol concentrations were significantly higher in LA/LB compared with HA/LB at the 30th and 45th minute of exercise (group–time interaction: $P = 0.02$). During exercise, AUC for glycerol was higher in LA/LB than that in HA/LB women ($P = 0.0005$) (Fig. 3D). No significant correlation was found between glycerol concentrations and A/LB FM ratio ($r = -0.25$, $P = 0.29$).

GH and ANP responses. Under resting conditions, plasma GH concentrations were not significantly different between the two groups ($P = 0.37$) (Fig. 4A). A group effect was observed for GH concentrations with greater plasma values in LA/LB when compared with HA/LB ($P = 0.03$) (Fig. 4A). The exercise induced an increase in plasma GH concentrations in all subjects (time effect: $P = 0.005$). During exercise, AUC for GH was lower in HA/LB than that in LA/LB women ($P = 0.003$) (Fig. 4B).

A group effect was observed for ANP concentrations with greater values in LA/LB than HA/LB women at rest ($P = 0.01$) and during exercise ($P = 0.009$) (Fig. 4C). During exercise, AUC also showed a group effect with greater ANP values in the LA/LB group ($P = 0.0007$) (Fig. 4D). No significant correlation was observed for GH ($r = -0.20$, $P = 0.40$) and ANP ($r = -0.27$, $P = 0.24$) concentrations and A/LB FM ratio.

Adrenaline and noradrenaline responses. Plasma adrenaline and noradrenaline concentrations were not different between groups at 12:00 p.m. ($P = 0.89$ and $P = 0.76$ for adrenaline and noradrenaline, respectively). Catecholamine responses did not differ between the two groups during exercise ($P = 0.91$ and $P = 0.88$ for adrenaline and

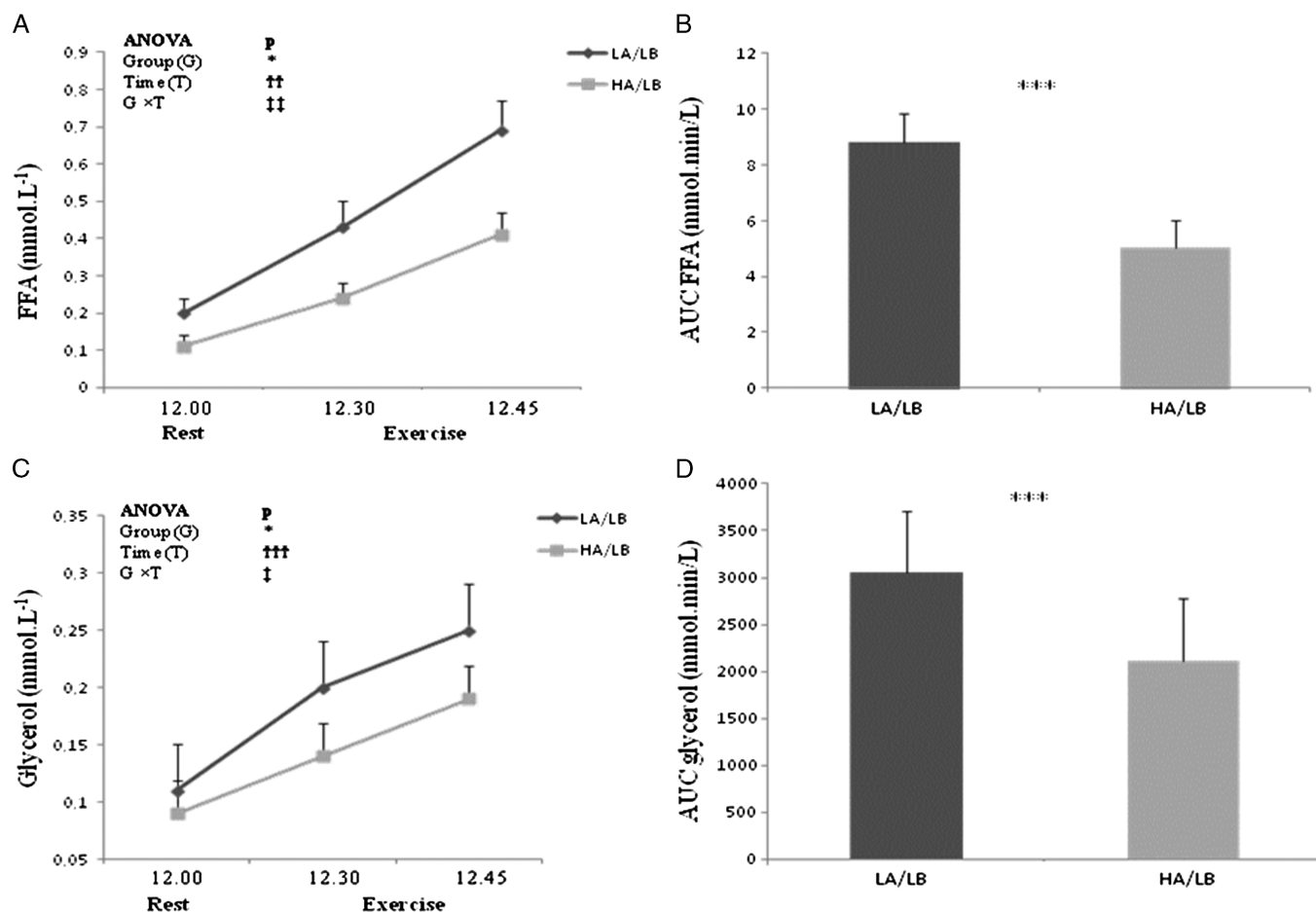


FIGURE 3—FFA and glycerol responses at rest and during exercise in both groups. **A**, FFA responses at rest and during exercise in both groups. **B**, AUC for FFA in both groups. **C**, Glycerol responses at rest and during exercise in both groups. **D**, AUC for glycerol in both groups. Data are presented as mean \pm SEM. Group effect, * $P < 0.05$ and *** $P < 0.001$; time effect, †† $P < 0.01$ and ††† $P < 0.001$; interaction group \times time, ‡ $P < 0.05$ and ‡‡ $P < 0.01$. LA/LB, low abdominal to lower body FM ratio group; HA/LB, high abdominal to lower body FM ratio group.

noradrenaline, respectively). A time effect was observed for catecholamines with greater values at the end of exercise (12:45 p.m.) compared with resting values ($P = 0.0005$ and $P < 10^{-6}$ for adrenaline and noradrenaline, respectively). No significant difference was observed between groups for AUC during exercise ($P = 0.90$ and $P = 0.65$ for adrenaline and noradrenaline, respectively), and there was no significant correlation between catecholamine ($r = 0.11$, $P = 0.66$; $r = 0.06$, $P = 0.80$ for adrenaline and noradrenaline, respectively) concentrations and A/LB FM ratio.

DISCUSSION

The present data indicated that A/LB FM ratio alters fuel use and metabolic and hormonal responses during exercise in premenopausal women with normal weight and WC. In this population, a lower A/LB FM ratio was associated with higher lipid mobilization and oxidation and a greater metabolic flexibility during exercise.

General characteristics. In the present study, $\dot{V}O_{2\max}$, total FM, total, and localized FFM values were not significantly different between the two groups. However, they

differed in terms of WC and A/LB FM ratio. As in normal weight women (20), normal body and FM values were associated with healthy metabolic profiles. To date, most studies showing that abdominal FM expansion is associated with insulin resistance and dyslipidemia whereas lower body FM is presented as a protective factor have been performed in subjects with obesity or type 2 diabetes (1,25). The present results showed that in normal weight individuals without HTGW or insulin resistance, there are no significant differences between the HA/LB and the LA/LB groups for fasting glucose and insulin concentrations, G/I, and HOMA-IR. However, measurements performed 3 h after a standardized breakfast showed that if plasma glucose concentrations still did not differ between both groups, insulin levels were significantly higher in the HA/LB group ($P < 0.01$). This suggests lower insulin sensitivity in the postprandial state in normal weight women with greater A/LB FM ratio, similarly to what is observed in overweight and obese women.

Abdominal FM in women has been shown to be associated with high plasma androgen levels (6), especially in conditions such as polycystic ovarian syndrome where women often present hyperandrogenism combined with

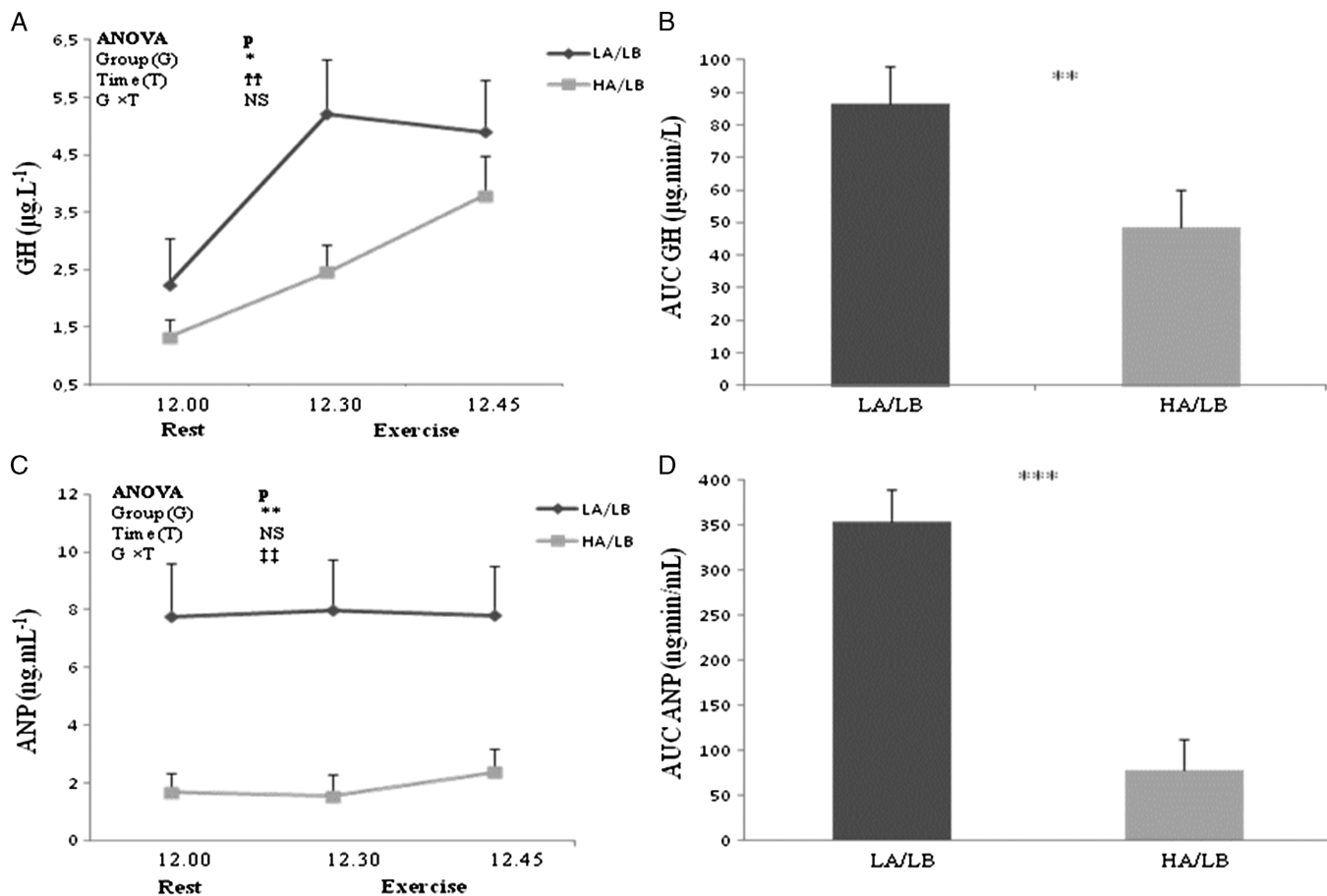


FIGURE 4—GH and ANP responses at rest and during exercise in both groups. A, GH responses at rest and during exercise in both groups. B, AUC for GH in both groups. C, ANP responses at rest and during exercise in both groups. D, AUC for ANP in both groups. Data are presented as mean \pm SEM. NS, not statistically significant; group effect, $*P < 0.05$, $**P < 0.01$, and $***P < 0.01$; time effect, $\dagger\dagger P < 0.01$; interaction group \times time, $\dagger\dagger\dagger P < 0.01$. LA/LB, low abdominal to lower body FM ratio group; HA/LB, high abdominal to lower body FM ratio group.

abdominal obesity, hyperinsulinemia, and oligomenorrhea (11). In the present study, however, we did not observe any effect of a higher A/LB FM ratio on the androgenic profile of premenopausal women with normal weight and WC.

Metabolic and hormonal responses to exercise.

Although there was no difference between the two groups at rest for substrate oxidation variables, women in the LA/LB group exhibited higher plasma FFA and glycerol levels indicating greater lipid mobilization as well as higher lipid use and contribution to energy expenditure during exercise. Although we did not assess the depletion of substrate pools such as glycogen and intramuscular TG, which contribute significantly to energy expenditure during exercise at 65% of $\dot{V}O_{2\max}$ (32), we could establish that whole-body substrate oxidation is affected by the adipose tissue localization, independently of FFM localization. This suggests that women with greater A/LB FM ratio exhibited a reduced ability to switch efficiently to a greater reliance on lipid oxidation during prolonged exercise, that is, altered metabolic flexibility (10). The negative correlation between A/LB fat mass ratio and lipid oxidation rates ($P < 0.05$) and the specific metabolic and hormonal responses to exercise further confirm that increased abdominal fat storage altered

lipid metabolism in premenopausal women with normal weight and WC. Thus, a higher A/LB FM ratio in normal weight women could be associated with low lipid turnover during exercise and may favor a risk of lipotoxicity and insulin resistance with age advancing.

There are, to our knowledge, little data available looking at the effect of preferential fat deposition at the abdominal or peripheral level on fuel metabolism in premenopausal normal weight women with normal WC. Toth et al. (36) observed lower lipid oxidation rate in postmenopausal compared with premenopausal women, which was in part explained by increased abdominal FM. Using the waist-to-hip ratio to assess adipose tissue localization, Kanaley et al. (18) observed that women with lower body obesity had greater plasma FFA responses but similar total lipid oxidation when compared with women with upper body obesity. In contrast, in our study, both plasma FFA and glycerol concentrations and lipid oxidation rates were increased in the LA/LB group compared with the HA/LB group. Furthermore, a negative correlation was found between A/LB FM ratio and FFA concentrations, indicating that, in addition to reduced ability for lipid oxidation, a high A/LB FM ratio in normal weight women is associated with low lipolytic ability.

However, this is inconsistent with previous studies showing that under resting condition, excess abdominal FM is rather characterized by greater lipolytic activity with lower glucose disposal and oxidation (23). At rest, Okura et al. (30) reported in overweight women that an excess in intra-abdominal FM resulted in an increase of lipolytic activity of intra-abdominal FM tissue and chronic excessive FFA release into the circulation leading to metabolic abnormalities. In our study, the transient increase in lipolytic activity induced by exercise, as opposed to chronically increased lipolysis with obesity, is likely to explain the lack of metabolic disturbance with high FFA plasma levels in our population. Considering the role of lipolytic and antilipolytic hormones (and adipocyte sensitivity) as factors determining lipid availability and use, we investigated hormonal responses to exercise (12).

Significantly higher postprandial plasma insulin levels in the HA/LB group, despite unaltered plasma glucose concentrations, indicated lower insulin sensitivity with preferential abdominal fat deposition. Insulin inhibits lipolysis and FFA flux and stimulates glucose uptake (21). Lipolysis inhibition and increased glucose uptake during exercise, caused by higher insulin levels in the HA/LB group, may explain lower lipid mobilization and contribution of lipid to energy expenditure compared with the LA/LB group. Difference in lipid metabolism between the two groups may in part be linked to differences in lipolytic stimulation by GH and ANP levels, which increased significantly more in response to exercise in the LA/LB group than that in the HA/LB. ANP is released from the heart and exerts its lipolytic effect through an increase in intracellular cGMP concentrations (21), a signaling pathway independent of insulin and catecholamines (mediated by cAMP). However, as GH response requires 2–3 h to increase lipid mobilization, its lipolytic effect was most likely negligible during the 45-min exercise performed in the present study (26,28). Adrenaline and noradrenaline are other potent lipolytic hormones acting through β_1 , β_2 , and β_3 adrenoreceptors whereas they inhibit lipolysis through α_2 adrenoreceptors (16,27). In resting conditions, intra-abdominal adipocytes are more sensitive to β -adrenergic stimulation and less sensitive to α -adrenergic receptors than subcutaneous lower body FM tissue (3). Contrary to ANP and GH, we did not observe any difference in catecholamine levels between the LA/LB and the HA/LB groups, although their level increased during exercise. Hence, this does not support a role for catecholamines in explaining differences for lipid metabolism between the LA/LB group and the HA/LB group. It can be first hypothesized that β -adrenergic sensitivity

is increased in all adipose tissues during exercise (21) or that cycling only provides a lipolytic stimulus localized to the lower limbs with little effect on lipid mobilization in abdominal adipose tissue. Investigation during arm cranking exercise or running in the LA/LB and the HA/LB groups could allow determining the effects of different muscle groups on lipid metabolism.

CONCLUSIONS

Even if further studies with greater sample size are needed to corroborate the present results, our study showed that using the A/LB FM ratio to assess preferential fat deposition allowed detecting altered whole-body lipid metabolism and hormonal responses during exercise in premenopausal women with normal weight, WC, and similar A/LB FFM ratio. Increased plasma FFA and glycerol levels and higher lipid oxidation rate in the LA/LB group indicated a greater ability for lipid mobilization and use in women with preferential lower body fat deposition during a 45-min cycling exercise. Lower insulin levels and higher ANP concentrations in this group of women are the factors most likely to explain better lipid mobilization when compared with women with higher abdominal fat deposition. Moreover, a lower metabolic flexibility in HA/LB women during exercise may place them at greater metabolic risks with age advancing.

The effect of adipose tissue deposition should be taken into account when physical activity is prescribed with the purpose of increasing energy expenditure, fat use, and maintaining adequate level of body fat. Similar lipid oxidation and level of adipose tissue lipolysis may be reached with exercise differing in intensity, duration, and modalities in LA/LB and HA/LB women. This may be of particular interest both in rehabilitation programs for athlete women and in physical activity prescriptions in recreationally active women.

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There is no conflict of interest.

The results of the present study do not constitute endorsement by the American College of Sports Medicine.

REFERENCES

1. Aasen G, Fagertun H, Tonstad S, Halse J. Leg fat mass as measured by dual x-ray absorptiometry (DXA) impacts insulin resistance differently in obese women versus men. *Scand J Clin Lab Invest*. 2009;69(2):181–9.
2. Alberti KG, Zimmet P, Shaw J. Metabolic syndrome—a new worldwide definition. A consensus statement from the International Diabetes Federation. *Diabet Med*. 2006;23(5):469–80.
3. Arner P, Hellstrom L, Wahrenberg H, Bronnegard M. Beta-adrenoceptor expression in human fat cells from different regions. *J Clin Invest*. 1990;86(5):1595–600.
4. Aucouturier J, Meyer M, Thivel D, Taillardat M, Duche P. Effect of android to gynoid fat ratio on insulin resistance in obese youth. *Arch Pediatr Adolesc Med*. 2009;163(9):826–31.

5. Blaak E. Gender differences in fat metabolism. *Curr Opin Clin Nutr Metab Care*. 2001;4(6):499–502.
6. Blouin K, Boivin A, Tchernof A. Androgens and body fat distribution. *J Steroid Biochem Mol Biol*. 2008;108(3–5):272–80.
7. Carter SL, Rennie C, Tarnopolsky MA. Substrate utilization during endurance exercise in men and women after endurance training. *Am J Physiol Endocrinol Metab*. 2001;280(6):E898–907.
8. Colberg SR, Simoneau JA, Thaete FL, Kelley DE. Skeletal muscle utilization of free fatty acids in women with visceral obesity. *J Clin Invest*. 1995;95(4):1846–53.
9. Despres JP, Lemieux I. Abdominal obesity and metabolic syndrome. *Nature*. 2006;444(7121):881–7.
10. DiPietro L. Exercise training and fat metabolism after menopause: implications for improved metabolic flexibility in aging. *J Appl Physiol*. 2010;109(6):1569–70.
11. Dunaif A. Insulin resistance and the polycystic ovary syndrome: mechanism and implications for pathogenesis. *Endocr Rev*. 1997;18(6):774–800.
12. Friedmann B, Kindermann W. Energy metabolism and regulatory hormones in women and men during endurance exercise. *Eur J Appl Physiol Occup Physiol*. 1989;59(1–2):1–9.
13. Glickman SG, Marn CS, Supiano MA, Dengel DR. Validity and reliability of dual-energy x-ray absorptiometry for the assessment of abdominal adiposity. *J Appl Physiol*. 2004;97(2):509–14.
14. Hamadeh MJ, Devries MC, Tarnopolsky MA. Estrogen supplementation reduces whole body leucine and carbohydrate oxidation and increases lipid oxidation in men during endurance exercise. *J Clin Endocrinol Metab*. 2005;90(6):3592–9.
15. Hatta H, Atomi Y, Shinohara S, Yamamoto Y, Yamada S. The effects of ovarian hormones on glucose and fatty acid oxidation during exercise in female ovariectomized rats. *Horm Metab Res*. 1988;20(10):609–11.
16. Ibrahim MM. Subcutaneous and visceral adipose tissue: structural and functional differences. *Obes Rev*. 2010;11(1):11–8.
17. Johnson JA, Fried SK, Pi-Sunyer FX, Albu JB. Impaired insulin action in subcutaneous adipocytes from women with visceral obesity. *Am J Physiol Endocrinol Metab*. 2001;280(1):E40–9.
18. Kanaley JA, Cryer PE, Jensen MD. Fatty acid kinetic responses to exercise. Effects of obesity, body fat distribution, and energy-restricted diet. *J Clin Invest*. 1993;92(1):255–61.
19. Koubi HE, Desplanches D, Gabrielle C, Cottet-Emard JM, Sempore B, Favier RJ. Exercise endurance and fuel utilization: a reevaluation of the effects of fasting. *J Appl Physiol*. 1991;70(3):1337–43.
20. Krotkiewski M, Seidell JC, Bjorntorp P. Glucose tolerance and hyperinsulinaemia in obese women: role of adipose tissue distribution, muscle fibre characteristics and androgens. *J Intern Med*. 1990;228(4):385–92.
21. Lafontan M, Moro C, Sengenès C, Galitzky J, Crampes F, Berlan M. An unsuspected metabolic role for atrial natriuretic peptides: the control of lipolysis, lipid mobilization, and systemic nonesterified fatty acids levels in humans. *Arterioscler Thromb Vasc Biol*. 2005;25(10):2032–42.
22. LaMonte MJ, Ainsworth BE, DuBose KD, et al. The hypertriglyceridemic waist phenotype among women. *Atherosclerosis*. 2003;171(1):123–30.
23. Lemieux S, Despres JP. Metabolic complications of visceral obesity: contribution to the aetiology of type 2 diabetes and implications for prevention and treatment. *Diabete Metab*. 1994;20(4):375–93.
24. Lemieux S, Prud'homme D, Bouchard C, Tremblay A, Despres JP. Sex differences in the relation of visceral adipose tissue accumulation to total body fatness. *Am J Clin Nutr*. 1993;58(4):463–7.
25. Manolopoulos KN, Karpe F, Frayn KN. Gluteofemoral body fat as a determinant of metabolic health. *Int J Obes (Lond)*. 2010;34(6):949–59.
26. Møller N, Gjedsted J, Gormsen L, Fuglsang J, Djurhuus C. Effects of growth hormone on lipid metabolism in humans. *Growth Horm IGF Res*. 2003;13(Suppl A):S18–21.
27. Mora-Rodriguez R, Hodgkinson BJ, Byerley LO, Coyle EF. Effects of beta-adrenergic receptor stimulation and blockade on substrate metabolism during submaximal exercise. *Am J Physiol Endocrinol Metab*. 2001;280(5):E752–60.
28. Norrelund H. The metabolic role of growth hormone in humans with particular reference to fasting. *Growth Horm IGF Res*. 2005;15(2):95–122.
29. Novotny R, Going S, Teegarden D, et al. Hispanic and Asian pubertal girls have higher android/gynoid fat ratio than whites. *Obesity (Silver Spring)*. 2007;15(6):1565–70.
30. Okura T, Nakata Y, Yamabuki K, Tanaka K. Regional body composition changes exhibit opposing effects on coronary heart disease risk factors. *Arterioscler Thromb Vasc Biol*. 2004;24(5):923–9.
31. Peronnet F, Massicotte D. Table of nonprotein respiratory quotient: an update. *Can J Sport Sci*. 1991;16(1):23–9.
32. Romijn JA, Coyle EF, Sidossis LS, et al. Regulation of endogenous fat and carbohydrate metabolism in relation to exercise intensity and duration. *Am J Physiol*. 1993;265(3 Pt 1):E380–91.
33. Sasai H, Katayama Y, Nakata Y, Ohkubo H, Tanaka K. Obesity phenotype and intra-abdominal fat responses to regular aerobic exercise. *Diabetes Res Clin Pract*. 2009;84(3):230–8.
34. Tarnopolsky MA. Gender differences in substrate metabolism during endurance exercise. *Can J Appl Physiol*. 2000;25(4):312–27.
35. Toth MJ, Gardner AW, Arciero PJ, Calles-Escandon J, Poehlman ET. Gender differences in fat oxidation and sympathetic nervous system activity at rest and during submaximal exercise in older individuals. *Clin Sci (Lond)*. 1998;95(1):59–66.
36. Toth MJ, Tchernof A, Sites CK, Poehlman ET. Menopause-related changes in body fat distribution. *Ann N Y Acad Sci*. 2000;904:502–6.
37. Tremblay J, Peronnet F, Massicotte D, Lavoie C. Carbohydrate supplementation and sex differences in fuel selection during exercise. *Med Sci Sports Exerc*. 2010;42(7):1314–23.
38. van Aggel-Leijssen DP, Saris WH, Wagenmakers AJ, Hul GB, van Baak MA. The effect of low-intensity exercise training on fat metabolism of obese women. *Obes Res*. 2001;9(2):86–96.
39. Van Beaumont W. Evaluation of hemoconcentration from hematocrit measurements. *J Appl Physiol*. 1972;32(5):712–3.
40. Williams PT. Relationship of adiposity to the population distribution of plasma triglyceride concentrations in vigorously active men and women. *Atherosclerosis*. 2004;174(2):363–71.