The effect of moderate consumption of non-nutritive sweeteners on glucose tolerance and body composition in rats

Ashley P. Tovar, James W. Navalta, Laura J. Kruskall, and John C. Young

Abstract: Glucose tolerance and body composition were determined in male rats given non-nutritive sweeteners (NNS) (aspartame or sucralose) in drinking water. Areas under the curve for glucose and insulin with NNS did not differ from control. NNS treatment had no effect on weight gain or percent body fat. Epididymal fat pad mass was higher with aspartame and the ratio of trunk to total fat was less with sucralose versus control, suggesting that NNS consumption altered body fat distribution.

Key words: body composition, dietary intake, carbohydrate metabolism.

Résumé : On évalue la tolérance au glucose et la composition corporelle chez des rats mâles ayant bu une eau additionnée d’un édulcorant non nutritif (NNS, aspartame ou sucralose). La surface sous la courbe (AUC) du glucose et de l’insuline dans la condition NNS ne diffère pas de la valeur du groupe de contrôle. Le traitement NNS n’a pas d’effet sur le gain de poids ou le pourcentage de gras corporel. Par rapport au groupe de contrôle, la masse du coussinet adipeux épidymaire est plus élevée dans le groupe aspartame et le ratio de la masse adipeuse trunc/total est plus faible dans le groupe sucralose, ce qui suggère que la consommation de NNS modifie la distribution des graisses corporelles.


Introduction

Non-nutritive sweeteners (NNS) have been introduced as replacements for sugar sweetening in beverages, reducing energy intake while maintaining sweetness. However, the consequences of this intervention on metabolic function are unclear. Although presumed to be metabolically inert, NNS consumption has been associated with weight gain and altered glucose tolerance in some human and animal studies (Fowler 2016), suggesting that NNS may, in fact, have metabolic effects including an altered ability to compensate for calories expected by the sweet taste. Aspartame, a methyl ester of aspartic acid and phenylalanine, and sucralose, a chlorinated derivative of sucrose, are widely used sweeteners in beverages and foods; their use has been linked to obesity, metabolic syndrome, and diabetes (Fowler 2016). The purpose of this study was to examine the effects of low-dose aspartame or sucrose treatment on weight gain, body composition, and glucose tolerance in rats where NNS delivery method and diet are consistent.

Methods

Male Sprague-Dawley rats (n = 30) (Taconic Biosciences, Rensselaer, N.Y.) were housed in pairs in the Laboratory Animal Care Facility on a 12 h light–dark cycle with ad libitum access to standard rat chow (PicoLab Rodent Diet 20; Lab Diet, St Louis, Mo.) and water. Animals were randomly assigned to 1 of 3 groups, aspartame (n = 10), sucralose (n = 10), or control (n = 10). Commercially available aspartame (Merisant, Chicago, Ill.) and sucralose (Heartland Food Products Group, Carmel, Ind.) were administered in drinking water. NNS doses were based on the average daily water intake of rats, 80–110 mL/(kg·day)–1 (Baker et al. 2006) and on the mean body weight of each NNS group. Doses for aspartame ranged from 6–8 mg/(kg·day)–1 in week 1 to 15–21 mg/(kg·day)–1 in week 6. Doses for sucralose ranged from 1.8–2.6 mg/(kg·day)–1 in week 1 to 4.9–6.7 mg/(kg·day)–1 in week 6. NNS doses were well below the reported toxic level of 4 g/(kg·day)–1 for aspartame (Magnuson et al. 2007) and 10 g/(kg·day)–1 for sucralose (Goldsmith 2000) and met the US Food and Drug Administration defined Acceptable Daily Intake of 50 mg/(kg body weight·day)–1 and 5 mg/(kg body weight·day)–1 for aspartame and sucralose, respectively (US FDA 2015). Initial body weight was not different between groups (P = 0.13); animals were weighed weekly to adjust dosage of NNS for growth. This study was reviewed and approved by the University of Nevada Institutional Animal Care and Use Committee (University of Nevada, Las Vegas, Nev.).

After 6 weeks of treatment, an oral glucose tolerance test (OGTT) was administered. Following an overnight fast (14 h), a blood sample for glucose and insulin was taken by tail clip prior to administering a 2 g/kg dose of a 50% weight–volume dextrose solution by gavage, and samples were taken at 15, 30, 60, and 120 min after dosing. Blood glucose was measured immediately by a glucose meter (Ascensia Contour, Bayer HealthCare, Indianapolis, Ind.) and test strips. Blood samples (300 μL) for insulin were collected into micro capillary tubes. Samples were centrifuged, and plasma was removed and stored at –70 °C for later analysis by radioimmunoassay (Millipore Corp, Billerica, Mass.). Following the OGTT, rats were euthanized and lean body mass and fat mass were determined by dual energy X-ray absorptiometry with small animal software (Lunar Prodigy, General Electric, Madison, Wisc.). The epididymal fat pads were then removed and weighed.

Statistical analysis was performed with Statistical Package for the Social Sciences (SPSS) 22.0 software (IBM Corp. Released 2013. Armonk, New York). One-way analysis of variance (ANOVA) and
posthoc Tukey’s tests were used to identify the differences in the dependent measures between groups. Significance was set to \( P < 0.05 \). Areas under the glucose and insulin response curves (AUC) were calculated via the trapezoid method. Data are reported by analyzing the mean ± SEM.

### Results

The response to an oral glucose load is shown in Fig. 1. Fasting glucose \( (P = 0.232) \) and insulin \( (P = 0.427) \) were not different between groups (Figs. 1A and 1B). No differences were found at any time point for the glucose or insulin response to the oral glucose load. Glucose AUC was not significantly different from control AUC for aspartame or sucralose groups \( (P = 0.28) \) (Fig. 1C). The insulin AUC was also not different for aspartame or sucralose groups versus control \( (P = 0.52) \) (Fig. 1D). The rate of weight gain was similar between the treatment groups and the control group, averaging 66 g/week for the first 3 weeks and 35 g/week for week 4. Total weight gain was 233 ± 10 g for the aspartame and 249 ± 5 g for the sucralose versus 239 ± 5 g for the control group \( (P = 0.27) \). Results of the body composition analysis are shown in Table 1. Percent body fat was not different between the aspartame group or the sucralose group and the control group \( (P = 0.24) \). However, the weight of the epididymal fat pads was 20% greater in the aspartame-treated rats than in the control animals \( (P = 0.042) \). No differences were found between either the aspartame group or the sucralose group and the control group for trunk fat \( (P = 0.32) \), total fat \( (P = 0.49) \), or the ratio of trunk fat to total fat \( (P = 0.07) \).

### Discussion

Although presumed to be metabolically inert, NNS consumption has been associated with weight gain and altered glucose tolerance in some human and animal studies (Fowler 2016), suggesting that NNS may, in fact, have metabolic effects. Possible mechanisms for the metabolic activity of NNS include altered ability to compensate for calories expected by sweet taste (Swithers et al. 2009; Davidson et al. 2011), alterations in the gut microbiota processing of nutrients (Palmas et al. 2014; Abou-Donia et al. 2008; Suez et al. 2014), and/or inhibition of intestinal alkaline phosphatase (Gul et al. 2017). Neither the area under the glucose response curve nor the area under the insulin response curve was different between aspartame treatment or sucralose treatment compared with control in response to an oral glucose load. Similarly, the rate of weight gain and the total weight gained was not different between either NNS treated group or the control group.

A hyperglycemic response to an oral glucose load was found when NNS were delivered in the diet (Swithers et al. 2012) or drinking water (Suez et al. 2014; Palmas et al. 2014; Mitsutomi et al. 2014; Gul et al. 2017) indicating a disconnect between the sweet taste and the learned response to control energy balance. However, when the glucose load was given by gavage bypassing the oral taste receptors, as in this study, a normal response to the glucose load was observed (Swithers et al. 2012). Conversely, glucose AUC was higher in response to an intraperitoneal glucose load in high-fat fed mice (Gul et al. 2017) and in mice with dietary-induced obesity (Mitsutomi et al. 2014) treated with aspartame.

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**Table 1. Animal characteristics.**

<table>
<thead>
<tr>
<th></th>
<th>Aspartame</th>
<th>Sucralose</th>
<th>Control</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial body weight (g)</td>
<td>147±2</td>
<td>153±3</td>
<td>152±1</td>
</tr>
<tr>
<td>Final body weight (g)</td>
<td>380±10</td>
<td>402±7</td>
<td>392±5</td>
</tr>
<tr>
<td>Lean mass (g)</td>
<td>303.6±8.6</td>
<td>325.5±6.9</td>
<td>316.4±6.4</td>
</tr>
<tr>
<td>Body fat %</td>
<td>19.3±0.70</td>
<td>17.7±0.67</td>
<td>17.6±0.91</td>
</tr>
<tr>
<td>Trunk fat (g)</td>
<td>40.9±2.9</td>
<td>35.4±1.5</td>
<td>41.8±4.4</td>
</tr>
<tr>
<td>Total fat (g)</td>
<td>73.1±3.9</td>
<td>69.5±2.6</td>
<td>67.2±3.3</td>
</tr>
<tr>
<td>Ratio trunk fat:total fat</td>
<td>0.547±0.037</td>
<td>0.487±0.006</td>
<td>0.602±0.044</td>
</tr>
<tr>
<td>Epididymal fat (g)</td>
<td>5.50±0.34*</td>
<td>5.00±0.24</td>
<td>4.55±0.19</td>
</tr>
<tr>
<td>Bone mineral density (g/cm²)</td>
<td>0.170±0.003</td>
<td>0.171±0.003</td>
<td>0.166±0.002</td>
</tr>
<tr>
<td>Fasting blood glucose (mmol/L)</td>
<td>2.93±0.16</td>
<td>3.27±0.11</td>
<td>3.02±0.15</td>
</tr>
<tr>
<td>Plasma insulin (pmol/L)</td>
<td>129.4±12.0</td>
<td>117.8±22.8</td>
<td>316.4±6.4</td>
</tr>
</tbody>
</table>

*Significantly different from control, \( P < 0.05 \). Values are mean ± SEM.
Palmas et al. (2014) found a higher glucose AUC in high-fat fed rats than in chow-fed rats, but no separate effect of aspartame on glucose AUC in either group; however, insulin-stimulated glucose disposal was impaired in both the aspartame treated groups. Similarly, NNS ingestion has been shown to alter the glyceremic response to an oral glucose load in high-fat fed rats (Suez et al. 2014) and in obese human subjects (Pepino et al. 2013), but not in healthy lean subjects (Ma et al. 2010). The finding of no effect of either aspartame or sucralose consumption on glucose tolerance in lean rats in this study is consistent with these findings. These results suggest that fat feeding and (or) obesity may be predisposing factors for the altered glucose response to NNS consumption.

Studies in which the animals did not have an alternative to the consumption of aspartame or sucralose reported a slower rate of growth or no difference in weight compared with control animals (Palmas et al. 2014; Mitsutomi et al. 2014; Goldsmith 2000; Beck et al. 2002) as was found in this study. Although the weight gain was the same for the aspartame and sucralose groups compared with the control group, an effect of NNS on body composition was noted. Epididymal fat pad mass was significantly increased in aspartame treated rats, as also reported in mice (Mitsutomi et al. 2014; Collison et al. 2013). Since epididymal fat pads may relate to visceral adiposity in humans (Gesta et al. 2006), these results suggest that NNS may induce a change in body composition. Conversely, the ratio of trunk fat to total fat tended lower in the sucralose-treated animals compared with the control animals suggesting a relative reduction in abdominal obesity. Together, these findings suggest that changes in body composition may be related to the specific NNS consumed.

In summary, the results of this study showed no adverse effect of low-dose aspartame or sucralose consumption on glucose metabolism. While total weight gain and percent body fat were not different between either NNS group and the control group, an increase in abdominal fat was found with aspartame whereas the ratio of trunk to total fat trended lower with sucralose. These results are consistent with the findings of metabolic effects of NNS independent of weight gain.

Conflict of interest statement

The authors report no conflicts of interest associated with this manuscript.

References


